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## Research Paper

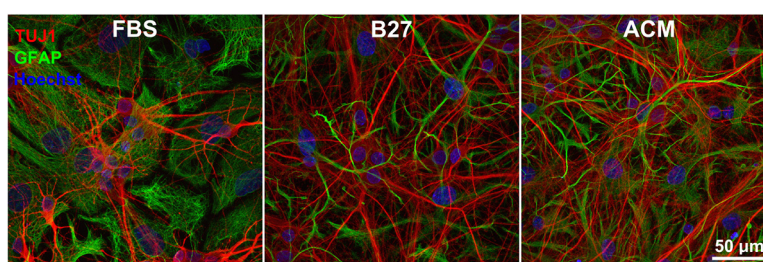
## An improved method for growing neurons: Comparison with standard protocols

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## HIGHLIGHTS

- An improved protocol for primary hippocampal cell cultures is proposed.
- The method relies on serum-free astrocytes conditioned medium (ACM).
- The ACM method is extensively compared with other two commonly used protocols.
- ACM improved morphology and function of both short- and long-term cultures.

## GRAPHICAL ABSTRACT



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## ABSTRACT

**Background:** Since different culturing parameters – such as media composition or cell density – lead to different experimental results, it is important to define the protocol used for neuronal cultures. The vital role of astrocytes in maintaining homeostasis of neurons – both *in vivo* and *in vitro* – is well established: the majority of improved culturing conditions for primary dissociated neuronal cultures rely on astrocytes. **New method:** Our culturing protocol is based on a novel serum-free preparation of astrocyte – conditioned medium (ACM). We compared the proposed ACM culturing method with other two commonly used methods Neurobasal/B27- and FBS- based media. We performed morphometric characterization by immunocytochemistry and functional analysis by calcium imaging for all three culture methods at 1, 7, 14 and 60 days *in vitro* (DIV).

**Results:** ACM-based cultures gave the best results for all tested criteria, *i.e.* growth cone's size and shape, neuronal outgrowth and branching, network activity and synchronization, maturation and long-term survival. The differences were more pronounced when compared with FBS-based medium. Neurobasal/B27 cultures were comparable to ACM for young cultures (DIV1), but not for culturing times longer than DIV7. **Comparison with existing method(s):** ACM-based cultures showed more robust neuronal outgrowth at DIV1. At DIV7 and 60, the activity of neuronal network grown in ACM had a more vigorous spontaneous electrical activity and a higher degree of synchronization.

**Conclusions:** We propose our ACM-based culture protocol as an improved and more suitable method for both short- and long-term neuronal cultures.

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## 1. Introduction

Primary dissociated neuronal cultures, obtained from embryonic or postnatal rodent brain regions such as the hippocampus or the cortex, represent a well-established *in vitro* model for studying neuronal networks, both on a short (network formation) and a long

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term (differentiation, maturation) scales (Dotti et al., 1988; da Silva and Dotti, 2002). When cells are dissociated, plated on appropriate substrates and cultured in appropriate media, they grow processes and form *ex-novo* a functional network.

Several media (N2, DMEM/F12) supplemented with serum, most commonly from fetal bovine (FBS) or horse origin, are commonly used for neuronal cultures. However, the chemical composition of the animal serum is not fully defined and includes some factors not present in the brain; moreover, the production of commercial serum is prone to batch to batch variability (Zheng et al., 2006; Arigony et al., 2013). For these reasons, the use of a chemically defined, serum-free medium is recommended for studies in which a complete control of the environment in the culture medium is required (Kivell et al., 2000). Several attempts were made in order to find serum free – media for optimal neuronal growth. Currently, the most widely used is Neurobasal medium with B27 supplement (Brewer et al., 1993; Beaudoin et al., 2012). Moreover, several modifications to Neurobasal/B27 were proposed with the addition/modification of glycoproteins, lipid and hormone molecules (NbActiv4, NS21) (Chen et al., 2008; Roth et al., 2010; Brewer et al., 2008). Most of those protocols were designed to promote long-term neuronal survival in an almost pure neuronal culture: neurons of embryonic (E18) and postnatal (P0) origin were cultured in the presence of <1% and 6–8% of glial cells, respectively (Beaudoin et al., 2012; Cullen et al., 2010).

However, the role of astrocytes for maintaining the homeostasis of neuronal cells both *in vivo* and *in vitro* has been clear for a long time (Araque and Navarrete, 2010; Nedergaard et al., 2003; Barker and Ullian, 2008; Allen and Barres, 2009; Banker, 1980). Different options are now available for a successful hippocampal cell culture, all of them involving astrocytes: I- direct plating of dissociated neurons on a glial feeder layer (Ivshitz and Segal, 2010), II- suspension of the coverslip with plated neurons above a glial feeder layer (Kaeck and Banker, 2006) and III- culturing neurons in astrocyte-conditioned medium (ACM). This third method has an advantage: it does not require a feeder cell's layer while providing all the soluble factors released by astrocytes, such as growth factors, signaling molecules and lipids (Hassanpoor et al., 2013; Ebrahimi et al., 2016; Mauch et al., 2001).

ACM has been demonstrated to increase neuronal survival and stem cell differentiation (Yamashita et al., 1992; Nakayama et al., 2003; Todd et al., 2013). However, the use of ACM in cell culturing protocols is still uncommon and often limited to *in vitro* models of pathologies, where its protective effect on neurons following mechanical injury and hypoxia has been demonstrated (Yan et al., 2013). Standard neuronal culturing methods are mostly based on serum-enriched or serum-free, supplement-enriched media (Beaudoin et al., 2012), both of which rely on commercially available resources. ACM, instead, can be obtained by the researcher from the same animal species used for primary neuronal cell's cultures.

Our goal is to establish an optimal, easy to use and reproducible culturing protocol for obtaining viable, functionally connected and long – term surviving primary hippocampal cultures. For this purpose, we compared two traditional culturing protocols with a new method based on the use of ACM as a growing medium. Our method differs from other previously published protocols (Todd et al., 2013) because the medium is produced in house and it is totally serum – free, unlike the commercial ACM obtained by conditioning of traditional astrocyte medium (supplemented with 10% FBS).

Our comparison is based on a morphometric and functional characterization of neuronal networks grown in the three different media. Above all, we describe a serum – free culturing method for long – term neuronal cultures with preserved functionality and connectivity, reminiscent of that seen in the intact tissue. The three culturing protocols that we compared were: I- FBS based

medium (traditionally used in our lab (Ulloa Severino et al., 2016) and by other research groups (Ivshitz and Segal, 2010; Cohen et al., 2008; Hughes et al., 2010); II- Neurobasal/B27 medium as described in Beaudoin (Beaudoin et al., 2012); III- our serum – free ACM medium. The exact composition for all the three growing media is reported in Section 5 and the conditions are referred to as FBS, B27 and ACM, respectively. The comparison between the three protocols was carried out by maintaining the same experimental parameters (cell density, animal age and functional tests). Therefore, the differences in morphology and electrical activity are attributed to the effect of the different media.

## 2. Results

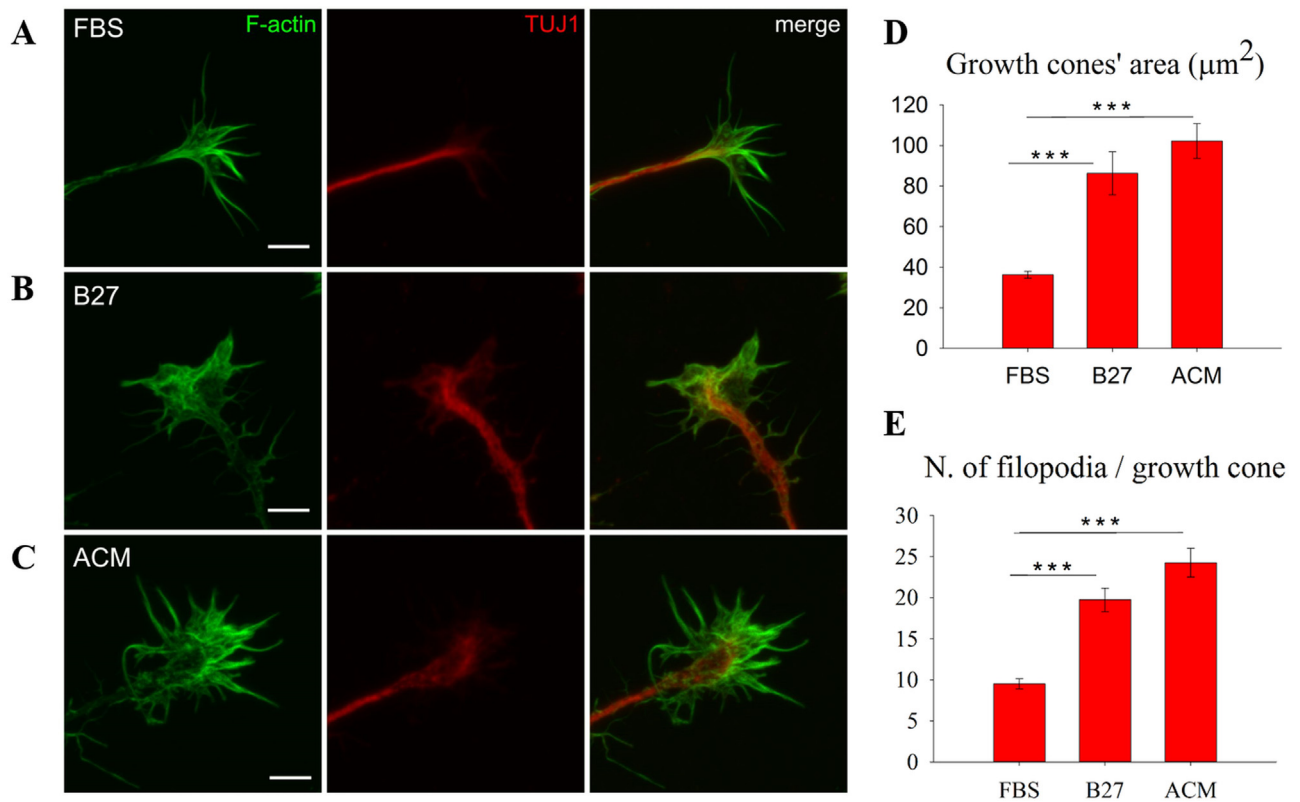
### 2.1. Setting of the plating protocol

We analyzed the morphology, electrical activity and long-term survival of our cell cultures grown in the three different media. All other parameters such as coverslip coating, plating medium and cells density were the same in the three cases. Briefly, after dissociation, the cells were suspended in plating medium (Neurobasal medium supplemented with 10% FBS, see Section 5 and Supplementary Table S1 for the complete formulation), seeded at a density of 100,000 cells/sample and incubated at 37 °C for half an hour in order to allow the cell's attachment on the coverslips previously coated with polyornithine (see Section 5). After that, the sample was divided in three groups – in this work referred to as FBS, B27 and ACM – and added to the appropriate growing medium. We used the same plating medium in the three cases because when cells were plated in serum-free Neurobasal/B27 medium, cells death increased and no electrical activity was detectable. Interestingly, when plated in ACM medium, the cells were able to survive for more than one month with consistent electrical activity (Supplementary Video S1).

### 2.2. Growth cones' morphology

Twenty-four hours after plating, we analyzed the cell morphology by immunocytochemistry. Cells were fixed and stained for filamentous actin (F-actin) and for  $\beta$ -tubulin III, a marker for post-mitotic neurons. At a first inspection, we observed a higher cell growth in Neurobasal/B27-based media, compared to FBS at DIV1. In particular, neuronal growth cones (GCs) were larger, thicker, richer in F-actin and had a more elaborated morphology with many filopodia (Fig. 1A–C). To quantify those differences, we measured and compared the surface of GCs among the three conditions. Confocal images were acquired with slice spacing of 0.2  $\mu\text{m}$ , up to 5  $\mu\text{m}$  z-stack thickness, and for each GC a maximum intensity projection of the stacks for F-actin was used. The contours of each GCs were manually traced and the surface was measured using ImageJ/Fiji. At least 20 GCs for each condition were analyzed (see Section 5). The GCs in Neurobasal/B27-based media were significantly larger comparing to FBS. GCs in ACM, in particular, showed the biggest size; however, the difference between ACM and B27 was not statistically significant ( $36.27 \pm 1.69 \mu\text{m}^2$  in FBS,  $86.27 \pm 10.63 \mu\text{m}^2$  in B27 and  $102.19 \pm 8.60 \mu\text{m}^2$  in ACM; Fig. 1D).

A similar analysis was done by counting the number of filopodia in each GC (Fig. 1F): GCs in FBS-based medium had on average  $9.48 \pm 0.62$  filopodia, B27  $19.72 \pm 1$  while ACM  $24.24 \pm 1.76$ , matching the most complex morphology observed in B27-based media (examples of elaborated forms – never observed in FBS – are shown in Supplementary Fig. S1).



**Fig. 1.** Neuronal growth cones at DIV1 for FBS (A), B27 (B) and ACM (C). Cells were stained for F-actin (left),  $\beta$ -tubulin III (TUJ1, middle) and merged (right). (D) Growth cone surfaces for the three conditions tested. (E) Number of filopodia per growth cone for FBS, B27 and ACM respectively. Scale bar, 5  $\mu\text{m}$ .  $n = 25$  growth cones for FBS,  $n = 29$  growth cones for B27,  $n = 25$  growth cones for ACM. \*\*\*  $p < 0.001$  One – Way ANOVA, Tukey post – hoc test.

**Table 1**

Number of neurites per cell at DIV1 in FBS, B27 and ACM represented as mean  $\pm$  standard error of the mean. Significantly higher values were observed in B27 and ACM comparing to FBS.

	Neurites/cell	Cells analyzed	Max number of neurites/cells
FBS	2,87 $\pm$ 0,10	205	9
B27	5,30 $\pm$ 0,18*	214	13
ACM	5,44 $\pm$ 0,22*	202	19

\*  $p < 0.001$ . One – Way ANOVA, Tukey post – hoc test.

### 2.3. Neurite outgrowth

Next, we analyzed the neurite outgrowth/branching. At DIV1, the overall morphology of neurons in Neurobasal/B27-based media was more elaborated and neurites in particular were thicker and had more spines (Fig. 2A–C). On the other hand, neurons in FBS had thinner neurites, simpler morphology, tended to grow straight and to cluster. To quantify the differences in the three conditions we considered, the same procedure was used: at DIV1, cells were fixed, stained and confocal images were acquired using a tubulin marker to calculate the average number of neurites per each neuron. FBS cultures had the lowest number ( $2.87 \pm 1.45$ ,  $n = 205$  neurons analyzed) comparing to the Neurobasal/B27-based media ( $5.30 \pm 2.61$  for B27 and  $5.44 \pm 3.06$  for ACM, respectively;  $n = 214$  neurons in B27 and  $n = 202$  neurons in ACM, Table 1). As for GCs, the ACM condition resulted in higher branching (up to 19 neurites per neuron, compared to maximal numbers of 9 and 13 for FBS and B27, respectively; Table 1) although not significantly different when compared to B27. These results confirmed that Neurobasal/B27-based media promote extensive neurite outgrowth resulting in higher branching of neurons.

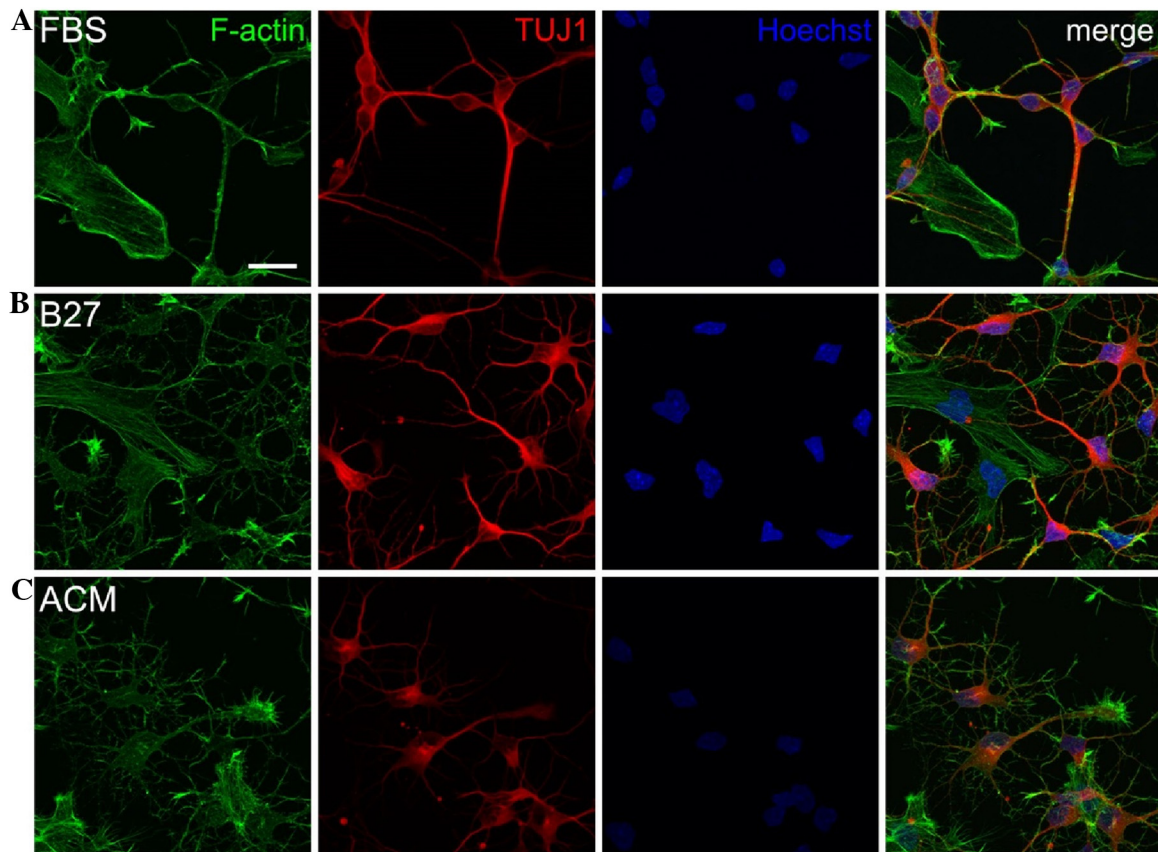
**Table 2**

Percentage of glia in the neuronal culture  $\pm$  standard error of the mean in the three conditions at DIV7, with the corresponding number of cells analyzed.

	% glia	N. cells analyzed
FBS	31,92 $\pm$ 2,48	704
B27	23,02 $\pm$ 2,02	648
ACM	29,16 $\pm$ 3,11	506

### 2.4. Astrocytes morphology and percentage

We did not observe significant differences in glial cells' morphology at this early developmental stage (DIV1, Supplementary Fig. S2). At DIV7, astrocytes grown in FBS and stained for glial fibrillary acidic protein (GFAP) assumed a flattened morphology (Fig. 3A). In Neurobasal/B27 media instead, complex shapes were observed with several cellular processes closely associated with neurons (Fig. 3B and C). Moreover, we compared the proportion of astrocytes between the three protocols. At DIV7, the percentage of glia in the neuronal cultures was estimated by counting GFAP – positive cells and observed to be around 32% in FBS, 23% in B27 and 29% in ACM (Table 2). As expected, because of the presence of several growth factors (Hassanpoor et al., 2013; Mauch et al., 2001), FBS and ACM resulted in higher percentages, despite the addition of the proliferation blocker cytosine- $\beta$ -D-arabino-furanoside (Ara-C) at DIV2 (see Section 5). The morphological differences were progressively more pronounced with the extension of the culturing time required for glial maturation. Astrocytes in ACM and B27 develop long and branched processes typical of a more mature morphology compared to the astrocytes in FBS at DIV14 (Supplementary Fig. S3).



**Fig. 2.** Neuronal branching at DIV1 for FBS (A), B27 (B) and ACM (C). From left to right: cells were stained for F-actin (green),  $\beta$ -tubulin III (TUJ1, red), Hoechst nuclear marker (blue) and then merged (right). Z-stacks of about  $10\ \mu\text{m}$  with  $0.2\ \mu\text{m}$  step size were acquired and maximum intensity projections of confocal images are here shown. Scale bar,  $20\ \mu\text{m}$ . (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

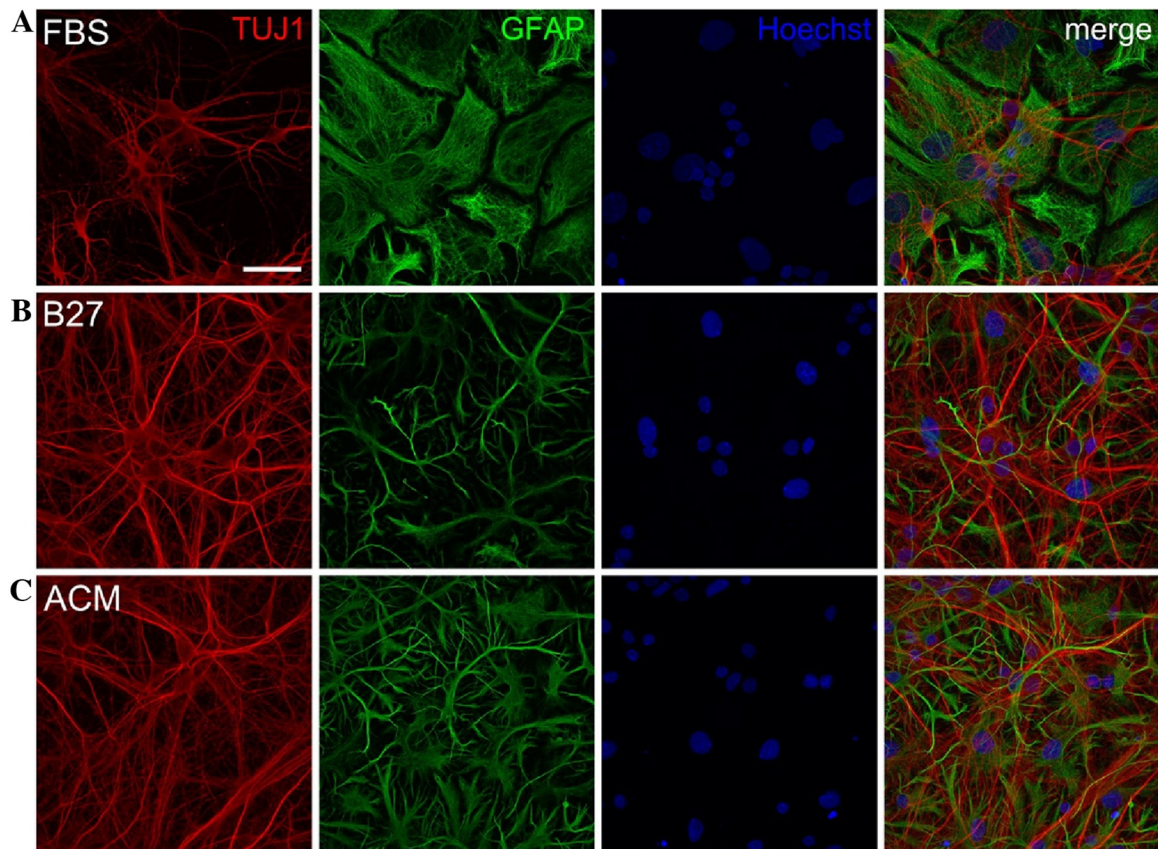
### 2.5. Functional properties of neuronal networks

Next we investigated the functional differences among neuronal networks. The spontaneous electrical activity of the neurons was analyzed by measuring their calcium transients ( $\text{DF}/\text{F}_0$ ) (Berridge, 1998), obtained by acquiring fluorescence images at 3–10 Hz for 10 min in every field of view. The onset time of calcium transients was defined by detecting those events in the fluorescence signal that exceeded at least by three times the standard deviation of the noise (about  $0.01\ \text{DF}/\text{F}_0$ , see Section 5). Moreover, we added tetrodotoxin (TTX) to the dish at the end of the recording (Supplementary Fig. S4) and verified the disappearance of calcium signals in the majority of cells in the culture, that were identified as neurons. The remaining cells were identified as astrocytes and excluded from the analysis, since they were showing calcium signals after TTX addition in the typical form of calcium waves (Scemes and Giaume, 2006).

Fluorescent images of the three cultures at DIV7, loaded with Oregon Green 488 BAPTA-1<sup>®</sup> cell-permeable calcium dye, and their representative traces are shown in Fig. 4A–C. The frequency of calcium peaks (Fig. 4D) showed an increasing trend from FBS ( $0.020 \pm 0.002$  peaks/s) to B27 ( $0.026 \pm 0.002$  peaks/s) and ACM ( $0.0322 \pm 0.0004$  peaks/s) with significant differences between FBS and ACM. Moreover, the mean time interval between calcium peaks (Inter Event Interval, IEI, Fig. 4E) was measured for every neuronal trace in order to analyze not only the frequency but also the distribution of peaks over time: the lowest value was observed in the ACM condition, with significant differences compared to FBS; an increase in the IEI was observed in B27 compared to ACM, although not significant ( $63.99 \pm 4.73$  s for FBS;  $57.06 \pm 9.53$  s for

B27;  $37.88 \pm 2.80$  s for ACM). Neurons grown in ACM showed a more homogeneous distribution of IEI comparing to the other conditions, as shown in Fig. 4G. Finally, the average correlation index (Fig. 4F) between the calcium peaks of different cells was calculated from cross correlation raster plots (see Section 5) and showed a significantly higher value in ACM ( $0.684 \pm 0.004$ ) when compared to both FBS ( $0.430 \pm 0.005$ ) and B27 ( $0.614 \pm 0.012$ ); a significantly higher value was also found in B27 compared to FBS ( $n = 157(5)$  couples of neurons (cultures) for FBS;  $n = 647(4)$  couples of neurons (cultures) for B27;  $n = 1680(4)$  couples of neurons (cultures) for ACM). Taken together, these data show that ACM cultures have higher frequency and synchronous activity compared to both FBS and B27.

Subsequently, we performed the same experiment with neuronal cultures at mature stages, *i.e.* 2 months growth (Fig. 5). The overall pattern of spontaneous electrical activity was found to be similar to the previously examined time point. The average frequency of calcium peaks (Fig. 5D) was still significantly increased in ACM when compared to the other two conditions, while a difference between B27 and FBS was no longer observed ( $0.063 \pm 0.006$  peaks/s in FBS;  $0.058 \pm 0.005$  peaks/s in B27;  $0.091 \pm 0.005$  peaks/s in ACM). An increased IEI (Fig. 5E) was found in FBS compared to both B27-based media ( $24.45 \pm 2.49$  s in FBS;  $23.03 \pm 2.67$  s in B27;  $17.35 \pm 1.45$  s in ACM). The distribution of IEI at this point was instead similar in the three conditions (Fig. 5G). Finally, a significantly higher cross correlation index (Fig. 5F) was measured in both B27-based media compared to FBS, and in ACM compared to B27 ( $0.62$  for  $363(4)$  couples of neurons (cultures) in FBS;  $0.77$  for  $504(4)$  couples of neurons (cultures) in B27;  $0.83$  for  $844(4)$  couples of neurons (cultures) in ACM). Thus, the neuronal cultures grown



**Fig. 3.** Neurons and glial cells morphology at DIV7 for FBS (A), B27 (B) and ACM (C). From left to right: cells were stained for  $\beta$ -tubulin III (TUJ1, red), GFAP (green), Hoechst nuclear staining (blue), and then merged. Confocal maximum intensity projections images of about 15  $\mu\text{m}$ -thick z stacks acquired with 0,5  $\mu\text{m}$  step size are here shown. Scale bar, 50  $\mu\text{m}$ . (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

in ACM maintain an overall higher spontaneous electrical activity and higher degree of synchrony over time compared to the cultures grown in traditional media. These results are consistent with previous works that demonstrate the advantage of using ACM for growing long – term cell cultures (Todd et al., 2013).

#### 2.6. Cell survival and morphology in the long – term cultures

Finally, we investigated the cell morphology of the three different cultures by immunocytochemistry. Neurons were stained with microtubule associated protein (MAP) 2 and astrocytes with GFAP (Fig. 6A–B). After 2 months, ACM cultures showed a denser neuronal network with higher cell survival, while in both FBS and B27 we observed a progressive increase of cell-free surface, most probably due to the loss of neurons. Astrocytes in FBS maintained a relatively simple, undifferentiated morphology with low degree of arborization. In contrast, GFAP staining in B27 revealed thin and long astrocytes processes. ACM cultures maintained a highly branched and complex phenotype, similar to mature astrocytes observed *in vivo* (Puschmann et al., 2013).

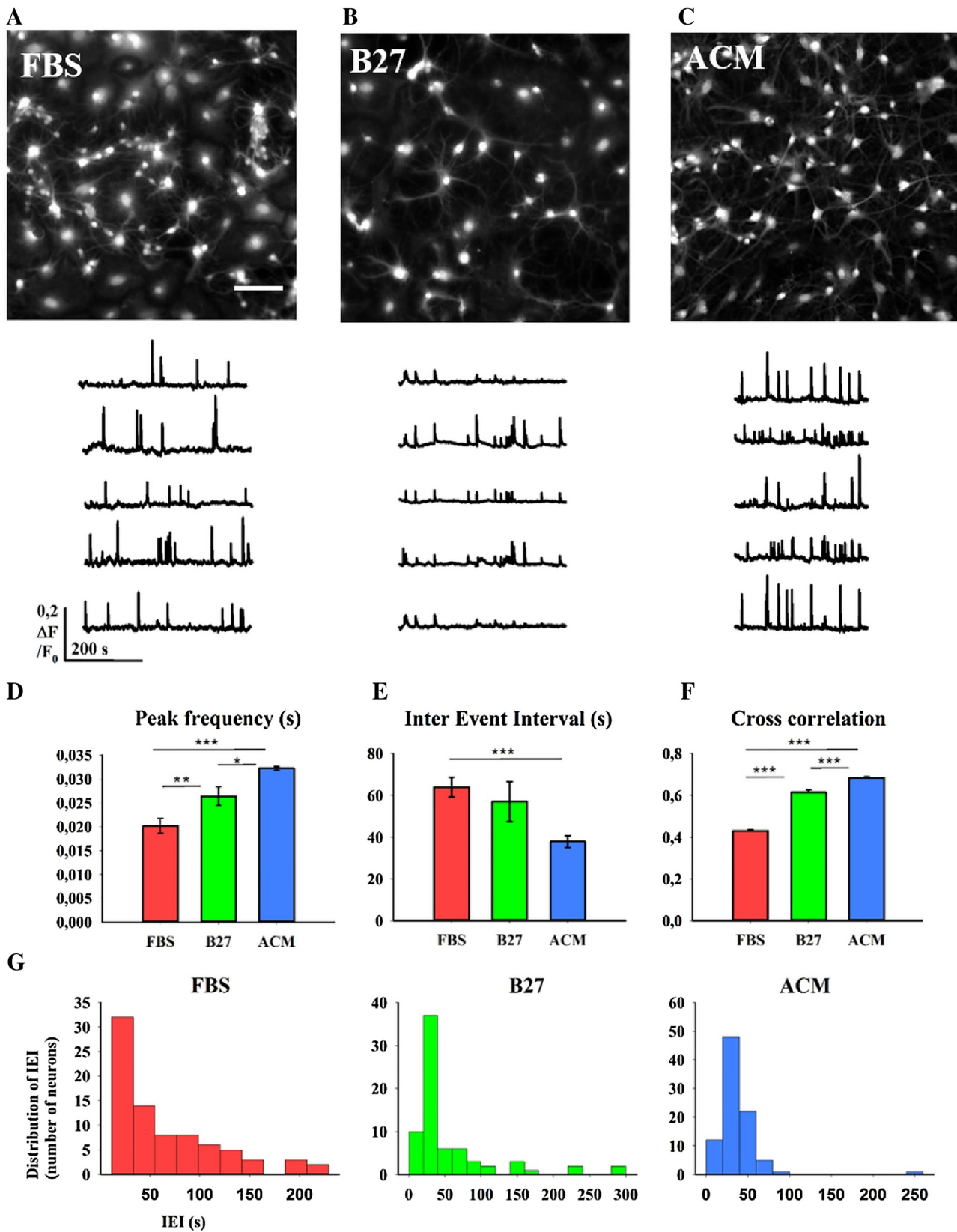
### 3. Discussion

Our aim is to establish a reproducible and easy – to – use protocol for growing long – term neuronal cultures with healthy and almost physiological functional properties. For this purpose, we produced a serum – free, Neurobasal/B27 – based ACM for growing primary hippocampal neurons directly after plating, unlike other methods in which neurons are first grown in serum – enriched media (Zhu et al., 2006). Unlike commercially available astrocyte – conditioned

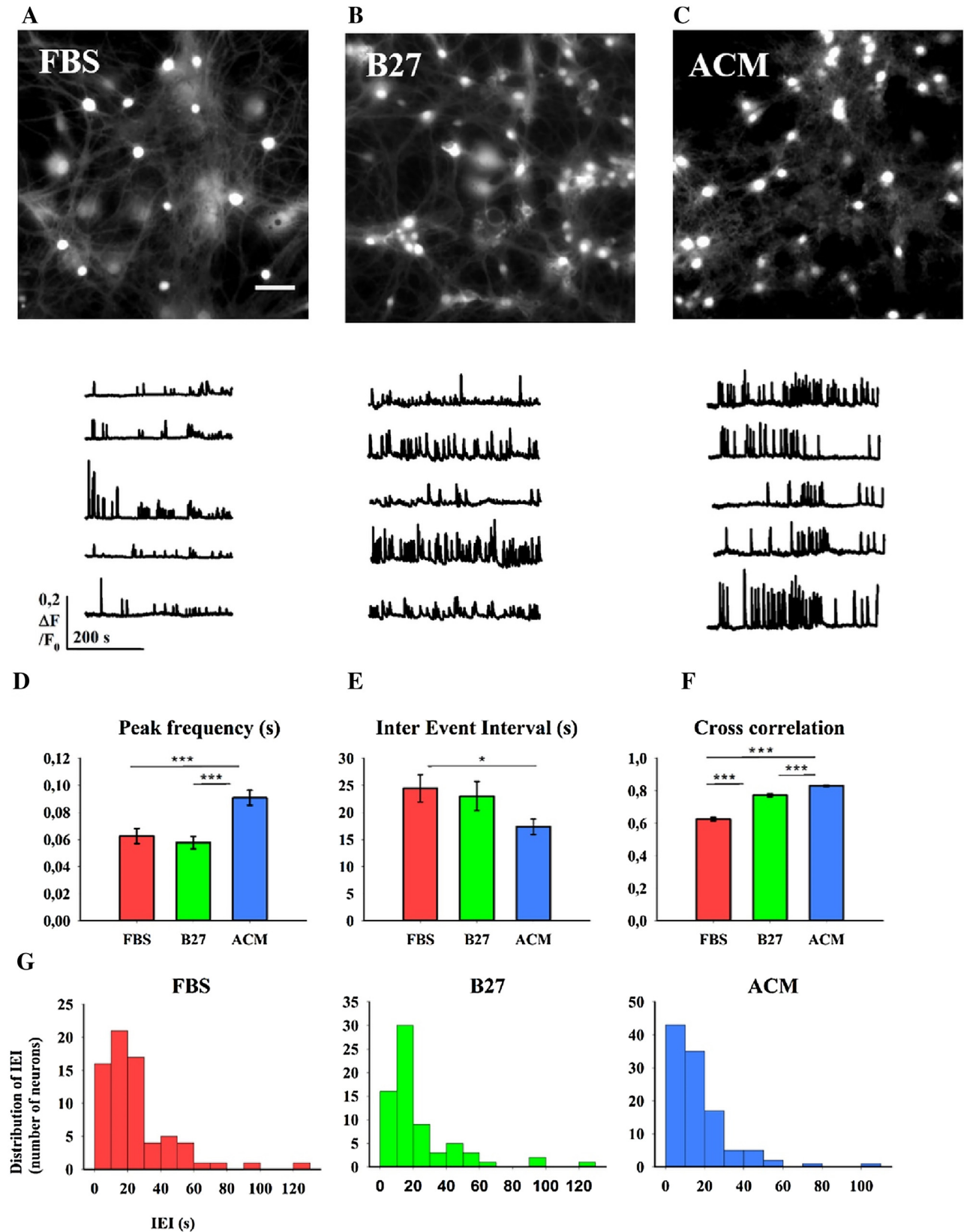
media, our ACM is derived from the same animal species and strain used for primary neuronal preparations and contains no additives. We harvested ACM as a growing medium, which allows a controlled percentage of glia in the culture. In this way, we preserved the physiological crosstalk between neurons and glia cells without using the glial feeder layer.

The resulting hippocampal cultures in ACM showed more robust neuronal outgrowth and branching compared to those grown in a serum – enriched medium, which makes them suitable for developmental studies as they provide an *in vitro* model reminiscent of the physiological CNS. Even though little is known about the *in vivo* morphology of axonal and dendritic growth cones, some works report evidence that the structure of neuronal growth cones is elaborated with numerous filopodia, especially at early developmental stages (Hossain et al., 2012). Astrocytes grown in serum – free conditions show a complex morphology that better represents *in vivo* systems (Puschmann et al., 2013; Wolfes et al., 2017) comparing to the flat astrocytic cell's bodies observed in FBS (compare astrocytes morphology between ACM and FBS in Fig. 3 and Supplementary Fig. S3). Indeed, the fibroblast-like morphology of astrocytes is known as an artifact due to the presence of serum in the culturing medium (Landis et al., 1990; Foo et al., 2011). Astrocytes in the hippocampus of adult rats show instead numerous branches and processes as previously demonstrated (Cerbai et al., 2012).

In addition to the improved short-term cultures, we obtained long-term neuronal cultures with preserved functional connectivity up to two months, as demonstrated by calcium imaging experiments. Different culturing conditions, i. e. physical properties and chemical composition of the medium used, may affect the affinity of the dye for calcium ions and the fluorescence intensity itself

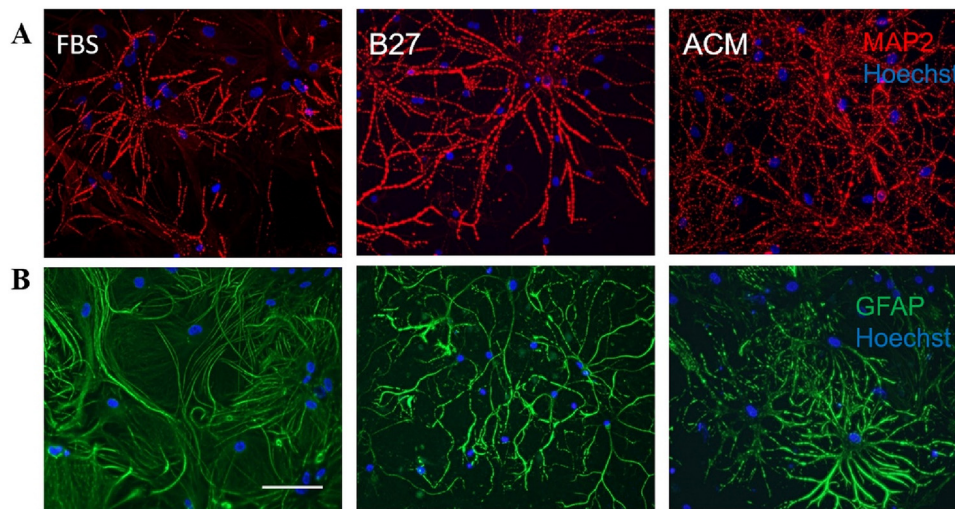


**Fig. 4.** Fluorescent images of neuronal cultures at DIV7 in FBS (A), B27 (B) and ACM (C) loaded with 4  $\mu$ M Oregon Green 488 BAPTA - 1<sup>o</sup> and corresponding calcium traces of 5 representative neurons simultaneously recorded. Average frequency (D), Inter Event Interval (E) and cross correlation index (F) of neuronal calcium peaks in the three conditions. Distribution of Inter Event Interval (IEI) in the three conditions (G). Scale bar, 100  $\mu$ m. n = 121 neurons for FBS; n = 71 neurons for B27; n = 98 neurons for ACM. \*p < 0.05 \*\*p < 0.01 \*\*\*p < 0.001 One-way ANOVA, Tukey post-hoc test. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)



**Fig. 5.** Fluorescent images of neuronal cultures grown for 2 months in FBS (A), B27 (B) and ACM (C) loaded with 4  $\mu$ M Oregon Green 488 BAPTA-1, AM<sup>®</sup> and corresponding calcium traces of 5 representative neurons simultaneously recorded. Average frequency (D), Inter Event Interval (E) and cross correlation index (F) of neuronal calcium peaks in the three conditions. Distribution of Inter Event Interval (IEI) in the three conditions (G). Scale bar, 100  $\mu$ m. n = 71 neurons for FBS; n = 70 neurons for B27; n = 110 neurons for ACM. \*p < 0.05 \*\*p < 0.001 One-way ANOVA, Tukey post-hoc test. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)





**Fig. 6.** Morphology of neurons (A) and astrocytes (B) grown for 2 months in FBS (left), B27 (middle) and ACM (right). Cells were stained for MAP2 (red), GFAP (green) and Hoechst nuclear staining (blue). Scale bar, 100  $\mu\text{m}$ . (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

(Oliver et al., 2000). In order to minimize these effects, we chose Oregon Green<sup>®</sup> 488 BAPTA-1, AM calcium dye because of its high-affinity and high signal-to-noise ratio (Grienberger and Konnerth, 2012). Moreover, the same experimental conditions such as dye concentration, temperature and incubation time were maintained for all the three protocols tested. A similar level of baseline fluorescence intensity was observed by visual inspection when recording cells grown in the three culturing media. Therefore, we assume that the uptake of the calcium sensitive dye from both neurons and astrocytes was similar in the three protocols, mainly dependent on the hydrophobicity of the AM – conjugated molecule (Paredes et al., 2008). However, further investigations are required to explain the role of astrocytes secreted factors in modulation of calcium-mediated neuronal processes (Bazargani and Attwell, 2016).

The presence of the differentiating factor B27 leads to an overall increase of the network's electrical activity when compared to what happens in the serum – based medium. However, the addition of ACM to the Neurobasal/B27 medium resulted in a further amplification of the overall spontaneous activity compared to both traditional methods (see Figs. 4 and 5). In particular, neurons grown in ACM showed significantly higher degree of correlation both in the short – and in the long – term compared to FBS and B27, suggesting that the synchronizing effect attributed to astrocytes may reside in their soluble factors (Amiri et al., 2012; Sasaki et al., 2014). Moreover, we demonstrated that ACM can totally replace the use of serum, even at the plating stage, while maintaining viability and functionality of the network (Supplementary Video S1). Although Neurobasal/B27 is the most commonly used serum-free medium for neuronal cultures (so called “maintenance medium” (Beaudoin et al., 2012)) it still requires the presence of FBS in the plating medium. Finally, since ACM alone is supposed to provide all the nutrients for long – term neuronal survival (e.g. lipids, BDNF and thrombospondins (Hassanpoor et al., 2013; Ebrahimi et al., 2016; Mauch et al., 2001)), the use of additional growth factors in the culturing medium is not necessary.

#### 4. Conclusions

We conclude that our ACM method is an improved and more suitable culture protocol for both short – and long – term studies, resulting in cell cultures that represent a realistic model of the CNS, useful for investigating the mechanisms of neuronal maturation and degeneration.

## 5. Materials and methods

### 5.1. Hippocampal cultures

Hippocampal neurons from Wistar rats (P2-P3) were prepared in accordance with the guidelines of the Italian Animal Welfare Act, and their use was approved by the Local Veterinary Service, the SISSA Ethics Committee board and the National Ministry of Health (Permit Number: 630-III/14) in accordance with the European Union guidelines for animal care (d.1.116/92; 86/609/C.E.). The animals were anaesthetized with CO<sub>2</sub> and sacrificed by decapitation, and all efforts were made to minimize suffering. Glass coverslips (15 mm diameter) were coated with 50  $\mu\text{g}/\text{ml}$  poly-L-ornithine (Sigma-Aldrich, St. Louis, MO, USA) overnight, and just before cells' seeding, a thin layer of Matrigel<sup>®</sup> (diluted 1:50 with culture medium; Corning, Tewksbury MA, USA) was applied. Dissociated cells were resuspended in Neuronal medium with the following composition: minimum essential medium (MEM) with GlutaMAX<sup>™</sup> supplemented with 10% dialyzed fetal bovine serum (FBS, all from Thermo Fisher Scientific, Waltham, MA, USA), 0.6% D-glucose, 15 mM Hepes, 0.1 mg/ml apo-transferrin, 30  $\mu\text{g}/\text{ml}$  insulin, 0.1  $\mu\text{g}/\text{ml}$  D-biotin, 1  $\mu\text{M}$  vitamin B12 and 2.5  $\mu\text{g}/\text{ml}$  gentamycin (all from Sigma-Aldrich). A drop containing 100,000 cells was deposited per each 15 mm diameter coverslip and incubated for 30 min at 37 °C. For each coverslip, placed in a 12-well tissue culture plate, 1.5 ml of culture medium was then added. Three sets of media were used: **I** – Neuronal medium with 10% FBS (the same used for resuspending cells, see above), **II** – Neurobasal medium supplemented with B27 and GlutaMAX<sup>™</sup> according to the manufacturer's instructions (all from Thermo Fisher) and 2.5  $\mu\text{g}/\text{ml}$  gentamycin, **III** – ACM (see below) in 1:1 ratio with Neurobasal/B27 medium. Half of the medium was changed after 48 h and 2  $\mu\text{M}$  cytosine- $\beta$ -D-arabinofuranoside (Ara-C; Sigma-Aldrich) was added to all culture media. The concentration of FBS (in medium containing 10% FBS) was decreased to 5%. Half of the medium was changed then twice per week. The neuronal cultures were maintained in an incubator at 37 °C, 5% CO<sub>2</sub> and 95% relative humidity. For a detailed description of the chemical composition of Neuronal vs Neurobasal medium, see Supplementary data.

## 5.2. Glia and ACM preparation

Glial cells were prepared as described in Kaech and Baker (Allen and Barres, 2009) with some modifications. Cortices from P2-P3 Wistar rats were dissociated as described for hippocampal cultures. Cells were plated in 15  $\mu\text{g/ml}$  poly-L-ornithine (Sigma-Aldrich) coated tissue-culture flasks at the density of  $7.5 \times 10^6$  cells per 75  $\text{cm}^2$  flask in Dulbecco's minimum essential medium (DMEM) with GlutaMAX™ (Thermo Fisher Scientific) supplemented with 10% fetal bovine serum (FBS) and Penicillin/Streptomycin (all from Euroclone, Pero, Italy). The next day the medium was changed in order to remove cell debris and loosely attached cells. Cells were cultured for ~1 week and medium was changed every 2–3 days. When almost confluent, cells were replated in 10 cm diameter Petri dishes at the density of  $5 \times 10^5$  cells per dish and expanded until they reached 75% confluence. The medium was then replaced with Neurobasal medium supplemented with B27 and GlutaMAX™ (all from ThermoFisher) and 2.5  $\mu\text{g/ml}$  gentamycin (Sigma-Aldrich). After 48 h ACM was collected, filtered and stored at 4 °C.

## 5.3. Immunocytochemistry and imaging

Cells were fixed in 4% paraformaldehyde containing 0.15% picric acid in phosphate-buffered saline (PBS), saturated with 0.1 M glycine, permeabilized with 0.1% Triton X-100, saturated with 0.5% BSA (all from Sigma-Aldrich) in PBS and then incubated for 1 h with primary mouse monoclonal antibodies: glial fibrillary acidic protein (GFAP, Sigma-Aldrich), anti- $\beta$ -tubulin III (TUJ1, Covance, Berkeley, CA) and mouse monoclonal anti-microtubule associated protein 2 (MAP2, Sigma-Aldrich). The secondary antibodies were goat anti-mouse Alexa Fluor® 594, goat anti-mouse immunoglobulin (Ig) G<sub>1</sub> Alexa Fluor® 488 and goat anti-mouse IgG<sub>2a</sub> Alexa Fluor® 594. F-actin was marked with Alexa Fluor 488® phalloidin (all from Thermo Fisher Scientific) and the incubation time was 30 min. Nuclei were stained with 2  $\mu\text{g/ml}$  in PBS Hoechst 33342 (Sigma-Aldrich) for 5 min. All the incubations were performed at room temperature (20–22 °C). The cells were examined using a Leica DMIRE2 confocal microscope (Leica Microsystems GmbH, Wetzlar, Germany) equipped with DIC and fluorescence optics, diode laser 405 nm, Ar/ArKr 488 nm and He/Ne 543/594 nm lasers. 63 $\times$  magnification and a 1.4 NA objective was used. Images were acquired at 1024  $\times$  1024 pixels resolution and z stacks with 200–250 nm steps. In addition, Leica DM6000 fluorescent microscope equipped with DIC and fluorescence optics, CCD camera and Volocity 5.4 3D imaging software (PerkinElmer, Coventry, UK) were used for cell counting experiments. The fluorescence images were collected with a 20 $\times$  magnification and 0.5 NA objective. For each image at least 30 slices were acquired with slice spacing of 0.5  $\mu\text{m}$ . Image J by W. Rasband (developed at the U.S. National Institutes of Health and available at <http://rsbweb.nih.gov/ij/>) was used for image processing.

## 5.4. Calcium imaging

The cells were incubated with the non-ratiometric calcium dye Oregon Green® 488 BAPTA-1, AM, (Thermo Fisher Scientific) dissolved in anhydrous DMSO (Sigma-Aldrich) at a concentration of 4 mM (stock solution), with the addition of Pluronic F-127 20% solution in dimethyl sulfoxide (DMSO, Thermo Fisher Scientific) for increasing the cell's permeability. The two components were dissolved at a ratio of 1:1 in Ringer's solution (145 mM NaCl, 3 mM KCl, 1.5 mM CaCl<sub>2</sub>, 1 mM MgCl<sub>2</sub>, 10 mM glucose and 10 mM Hepes, pH 7.4) to give a final dye concentration of 4  $\mu\text{M}$ , and incubated at 37 °C for 30 min. The cultures were then transferred to a glass-bottom Petri dish in order to allow visualization in a Nikon Eclipse Ti-U inverted microscope equipped with an HBO 103 W/2 mercury short

arc lamp (Osram, Munich, Germany), a mirror unit (exciter filter BP 465–495 nm, dichroic 505 nm, emission filter BP 515–555) and an Electron Multiplier CCD Camera C9100-13 (Hamamatsu Photonics, Japan). The experiments were performed at RT for 20 min/sample. At the end of each registration, the antagonist of voltage-gated sodium channels tetrodotoxin (TTX, Sigma-Aldrich) was added to the culture at a concentration of 1  $\mu\text{M}$  and the sample was recorded for additional 5 min.

Images were acquired using the NIS Element software (Nikon, Japan) with an S-Fluor 20 $\times$ /0.75 NA objective at a sampling rate of 3–10 Hz. The spatial resolution was 256  $\times$  256 pixels. To avoid saturation of the signals, excitation light intensity was attenuated by ND4 and ND8 neutral density filters (Nikon).

## 5.5. Data analysis

### 5.5.1. Morphometric analysis, neurite and cell counting

At least three different preparations were used. For neurite counting, nuclear stain was used to acquire randomly 3–5 fields with comparable cell densities (approx. 30 cells/field). TUJ1-stained neurites were counted and a mean number of neurites per neuron was calculated for all three tested conditions.

### 5.5.2. Ca<sup>2+</sup> imaging processing and analysis

As described in our previous work (Ulloa Severino et al., 2016; Moshtagh-Khorasani et al., 2013), the initial video was processed with the ImageJ (U. S. National Institutes of Health, Bethesda, MA) software. Briefly, neurons were localized by selecting the cells bodies with a region of interest (ROI), and an additional ROI was selected to subtract the background. The fluorescence intensity over time,  $I_f(t)$ , of every selected ROI was displayed and the extent of its decay – consequence of dye bleaching – was evaluated. To compensate photo bleaching, the decay of  $I_f(t)$  was fitted with a cubic or polynomial spline ( $Y(t)$ ) interpolating  $I_f(t)$  at 10 or 20 points. The function  $Y(t)$  fitting  $I_f(t)$  was then added to the original optical signal – to compensate dye bleaching – and the fractional optical signal was taken as  $DF/F_0 = Y(t) + I_f(t)/I_f(0)$ , where  $I_f(0)$  is the fluorescence intensity at the beginning of the recording.

### 5.5.3. Computation of raster plot and correlation coefficient of Ca<sup>2+</sup> transient occurrence

The times,  $t_i$ , at which transient peaks occurred are presented in a conventional raster plot.

The correlation coefficient of the calcium transients for neuron  $i$  and neuron  $j$  ( $s_{CTij}$ ) was computed as follows: the total recording time,  $T_{\text{tot}}$ , was divided into  $N$  intervals (1, ...,  $n$ , ...,  $N$ ) of a duration  $Dt$ , that was fixed at 20 s. Thus, if  $f_{in}$  and  $f_{jn}$  are the number of calcium transients of neuron  $i$  and neuron  $j$  in the time interval  $Dt_n$ , then

$$\sigma_{CTij} = \frac{\sum_n f_{in} f_{jn}}{\sqrt{(\sum_n f_{in}^2)(\sum_n f_{jn}^2)}}$$

so that  $s_{CTij}$  depends on  $Dt$  and varies between 0 and 1.

### 5.5.4. Statistical analysis

Data are shown as the mean  $\pm$  standard error of the mean (s.e.m.) from the number of samples indicated in each experiment (see Section 2). The mean values of both morphological and activity parameters were compared among the three different conditions with One-way ANOVA test followed by Tukey post-hoc comparison, using the software SigmaPlot 10.0. Significant differences were set with  $p$  values inferior to 0.05 (\* $p$  < 0.05, \*\* $p$  < 0.01, \*\*\* $p$  < 0.001).

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## Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.jneumeth.2017.01.013>.

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